

# Impaired Adaptive Cellular Responses to Oxidative Stress and the Pathogenesis of Alzheimer's Disease

Sarah J. Texel<sup>1,2</sup> and Mark P. Mattson<sup>1,2</sup>

## Abstract

As is generally true with other age-related diseases, Alzheimer's disease (AD) involves oxidative damage to cellular components in the affected tissue, in this case the brain. The causes and consequences of oxidative stress in neurons in AD are not fully understood, but considerable evidence points to important roles for accumulation of amyloid  $\beta$ -peptide upstream of oxidative stress and perturbed cellular  $\text{Ca}^{2+}$  homeostasis and energy metabolism downstream of oxidative stress. The identification of mutations in the  $\beta$ -amyloid precursor protein and presenilin-1 as causes of some cases of early onset inherited AD, and the development of cell culture and animal models based on these mutations has greatly enhanced our understanding of the AD process, and has greatly expanded opportunities for preclinical testing of potential therapeutic interventions. In this regard, and of particular interest to us, is the elucidation of adaptive cellular stress response pathways (ACSRP) that can counteract multiple steps in the AD neurodegenerative cascades, thereby limiting oxidative damage and preserving cognitive function. ACSRPs can be activated by factors ranging from exercise and dietary energy restriction, to drugs and phytochemicals. In this article we provide an overview of oxidative stress and AD, with a focus on ACSRPs and their potential for preventing and treating AD. *Antioxid. Redox Signal.* 14, 1519–1534.

## Introduction

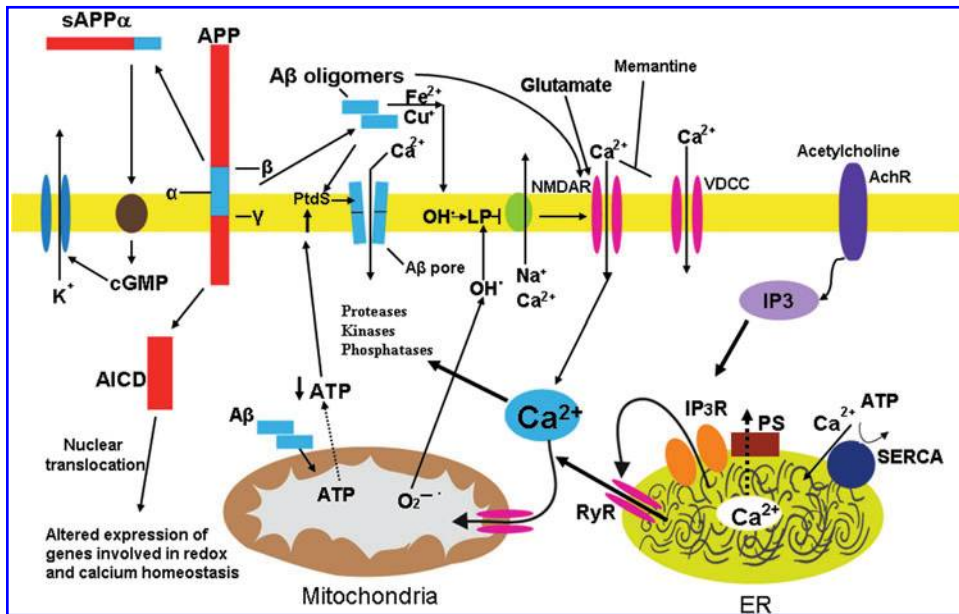
**A**LZHEIMER'S DISEASE (AD) was first described as a relatively rare disorder by Alois Alzheimer in 1906 and now, ~1 century later, nearly 5 million Americans are living with AD and the numbers are rapidly rising as baby boomers enter the AD danger zone of >65 years of age. Clinically, AD is diagnosed by (progressive) memory impairment and reduced size of the hippocampus, temporal, and frontal lobes as detected by magnetic resonance imaging analysis. One of the hallmark pathologies in AD is the altered proteolytic processing of the amyloid precursor protein (APP) that leads to accumulation of amyloid  $\beta$ -peptide ( $\text{A}\beta$ ) in extracellular plaques (112). Another prominent alteration is the presence of the so-called neurofibrillary tangles that are fibrillar bundles of the microtubule-associated protein tau A- $\beta$  plaques and tangles (17). There is abundant evidence that oxidative stress plays a role in nerve cell dysfunction and death in AD. Because this evidence has been reviewed previously (25, 117), we will briefly describe the salient features of the events that appear to play major roles in generating reactive oxygen species (ROS) in AD on the one hand, and the mechanisms by which ROS contribute to synaptic dysfunction and neuronal degeneration.

Large spherical (hundreds of micrometers in diameter) extracellular accumulations of  $\text{A}\beta$ , known as amyloid plaques, are a defining feature of AD. There is considerable evidence that  $\text{A}\beta$  can damage and kill neurons by a mechanism involving oxidative stress (Fig. 1). In AD,  $\text{A}\beta$  self-aggregates, and when small oligomers of  $\text{A}\beta$  are forming in the early stages of aggregation, hydrogen peroxide is generated from the peptide itself in a process requiring oxygen and trace amounts of  $\text{Fe}^{2+}$  and  $\text{Cu}^{+}$  (25).  $\text{A}\beta$  aggregation tends to occur on cell membranes resulting in membrane lipid peroxidation and the generation of the toxic aldehyde 4-hydroxynonenal, which can impair synaptic function and disrupt cellular  $\text{Ca}^{2+}$  and energy metabolism by covalently modifying proteins on cysteine, lysine, and histidine residues. Proteins whose functions have been shown to be impaired by 4-hydroxynonenal are plasma membrane  $\text{Na}^{+}/\text{K}^{+}$ - and  $\text{Ca}^{2+}$ -ATPases, and glucose and glutamate (glutamate is the major excitatory neurotransmitter in the brain) transporters (113). As a consequence, neurons become unable to maintain cellular ion homeostasis, energy is depleted, and the neurons may degenerate as the result of  $\text{Ca}^{2+}$  overload and triggering of apoptosis (Fig. 1).

Although  $\text{A}\beta$  plays an important role in oxidative stress and neuronal degeneration in AD, the presence of  $\text{A}\beta$  plaques

<sup>1</sup>Laboratory of Neurosciences, National Institute of Aging Intramural Research Program, Baltimore, Maryland.

<sup>2</sup>Department of Neuroscience, Johns Hopkins University School of Medicine, Baltimore, Maryland.



**FIG. 1. Molecular and cellular alterations involved in neuronal dysfunction in Alzheimer's disease (AD).** The  $\beta$ -amyloid precursor protein (APP) is cleaved by  $\beta$ -secretase ( $\beta$ ) and  $\gamma$ -secretase ( $\gamma$ ), resulting in the liberation of the amyloid  $\beta$ -peptide ( $A\beta$ ).  $A\beta$  may interact with  $Fe^{2+}$  and  $Cu^{2+}$  to generate hydrogen peroxide and hydroxyl radical, resulting in membrane lipid peroxidation that impairs the function of membrane  $Na^{+}$  and  $Ca^{2+}$  ATPases (ion pumps). The membrane then depolarizes, and glutamate receptor channels (N-methyl-D-aspartate receptor, N-methyl-D-aspartate receptor) and voltage-dependent  $Ca^{2+}$  channels (VDCC) open and  $Ca^{2+}$  enters the cyto-

plasm.  $A\beta$  may also form  $Ca^{2+}$ -permeable pores in the plasma membrane; the interaction of  $A\beta$  with the plasma membrane may be facilitated by binding to phosphatidylserine (PtdS).  $A\beta$  may also act directly on mitochondria to induce superoxide anion radical ( $O_2^{\bullet-}$ ) production,  $Ca^{2+}$  overload, and decreased ATP production. Amyloidogenic APP processing prevents  $\alpha$ -secretase ( $\alpha$ ) cleavage of APP, which normally generates an activity-dependent secreted form of APP (sAPP $\alpha$ ) that engages a signaling pathway involving cyclic guanosine monophosphate (cGMP) that activates  $K^{+}$  channels, thereby hyperpolarizing the membrane and reducing  $Ca^{2+}$  influx and free radical production. Amyloidogenic processing also generates an intracellular APP domain (AICD) that can translocate to the nucleus and modify gene transcription in ways that perturb redox and  $Ca^{2+}$  homeostasis. Presenilin-1 (PS) functions as a  $Ca^{2+}$  leak channel in the endoplasmic reticulum (ER) and PS mutations may impair this  $Ca^{2+}$  leak channel function resulting in excessive accumulation of  $Ca^{2+}$  in the ER and enhanced  $Ca^{2+}$  release through ryanodine receptor (RyR) and IP<sub>3</sub> receptor (IP<sub>3</sub>R) channels. There is also evidence that PS can interact directly or indirectly with RyR and smooth ER  $Ca^{2+}$ -ATPase (SERCA) to alter ER  $Ca^{2+}$  release and uptake. Altogether, the cascade of events described here first impairs synaptic transmission and may ultimately kill neurons in AD. Modified from Bezprozvanny and Mattson (16). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

is not sufficient for a diagnosis of AD. The reason is that some individuals with perfectly normal cognitive function exhibit large amounts of plaques (84), suggesting that neurons in these individuals are able to withstand any oxidative attack by  $A\beta$ . We are learning that there may be mechanisms by which brain cells respond adaptively to aging and counteract disease processes. This concept falls under the broader definition of hormesis, a process in which cells respond to low levels of stress by activating adaptive cellular stress response pathways (ACSRP) that promote cell repair and survival (110). In this situation hormesis acts to precondition cells, so they are better prepared when larger insults strike. Studies have shown that mental and physical activity can stimulate ACSRPs in the brain, which may be the reason physically and mentally active individuals are at a reduced risk for AD. On the other hand, factors such as obesity and diabetes may increase the risk for AD by impairing ACSRPs. The notion of failed ACSRPs as a pivotal factor in AD, and of lifelong activation of ACSRPs conferring resistance to AD, is the subject of the remainder of this article. While the present article focuses on adaptive cellular stress responses, there are many other molecular and cellular processes that are not stress responses, whose dysfunction has been implicated in the pathogenesis of AD, including the amyloid cascade,

oxidative stress, accumulation of damaged proteins, and mitochondrial impairment (112).

### Evidence That ACSRPs Are Impaired in AD

Studies of neurons in culture and *in vivo* have elucidated mechanisms by which the cells can increase their resistance to a range of adverse conditions, including oxidative stress. One mechanism involves the activity-dependent production of neurotrophic factors that activate receptors coupled to kinases and transcription factors that induce expression of genes encoding cytoprotective proteins. Three growth factors known to support the survival of neurons that are vulnerable in AD are fibroblast growth factor 2 (FGF2), nerve growth factor (NGF), and brain-derived neurotrophic factor (BDNF). FGF2 can protect neurons from the pathogenic actions of mutant presenilin-1 (PS1) and  $A\beta$ , and may do so by suppressing oxidative stress and stabilizing cellular calcium homeostasis (65). FGF2, NGF, and BDNF have all been shown to increase the resistance of neurons to oxidative stress, most likely by upregulating expression of antioxidant enzymes such as superoxide dismutases (SOD) and antiapoptotic proteins such as Bcl-2.

In AD, FGF2 may be sequestered in  $A\beta$  plaques, thereby reducing the amount of FGF2 available to activate its receptors

in neurons. NGF supports the survival and plasticity of cholinergic neurons in the basal forebrain; these neurons innervate the hippocampus where their axon terminals release acetylcholine, a process critical for learning and memory (172). The acetylcholinesterase inhibitors used to treat AD patients can enhance learning and memory by increasing the amount of synaptic acetylcholine. Studies of postmortem brain tissue from AD patients demonstrated a depletion of NGF in brain regions affected by the disease, including the hippocampus. The potential therapeutic benefit of NGF was tested in clinical trials in which NGF was infused into the lateral ventricle of AD patients; unfortunately, the NGF caused intolerable back pain, presumably because of actions in the spinal cord, and the trial was halted. BDNF is a particularly important neurotrophic factor because it is produced and released from neurons in an activity-dependent manner, and plays pivotal roles in synaptic plasticity, learning and memory, neuron survival, and neurogenesis (118, 156). Brain tissue samples from AD patients exhibit reduced levels of both BDNF and activated cyclic AMP response element-binding protein (CREB), a transcription factor that induces BDNF production (157). BDNF production is stimulated by at least three different behaviors that are believed to reduce the risk for AD, namely, exercise, cognitive stimulation, and dietary energy restriction (see next section below).

Three additional ACSRP that may be compromised in aging and AD are antioxidant response systems, protein chaperone systems, and protein degradation pathways. The increase in oxidative stress that occurs in brain cells with aging and early in the course of AD is associated with compensatory upregulation of some antioxidant enzymes (187), but also the impairment of other antioxidant defenses such as

the plasma membrane redox system (78), and depletion of low-molecular-weight antioxidants, including glutathione (102). Protein chaperones such as heat shock protein 70 (HSP70), glucose-regulated protein 78 (GRP78), and HSP27 have been reported to modify one or more processes involved in AD, including A $\beta$  aggregation, A $\beta$  toxicity, and oxidative stress (88). The accumulation of A $\beta$ , tau, and other proteotoxic proteins may normally be prevented, in part, by degradation of the aberrant proteins in the proteasome (Fig. 2). However, in AD the function of the proteasome is impaired, apparently as the result of oxidative damage to the proteasome proteins themselves (29, 85). So, it appears that beginning early in the disease process neurons in AD suffer from increased oxidative and proteotoxic stress, adaptive responses to this cellular stress are engaged, but ultimately the ACSRPs fail.

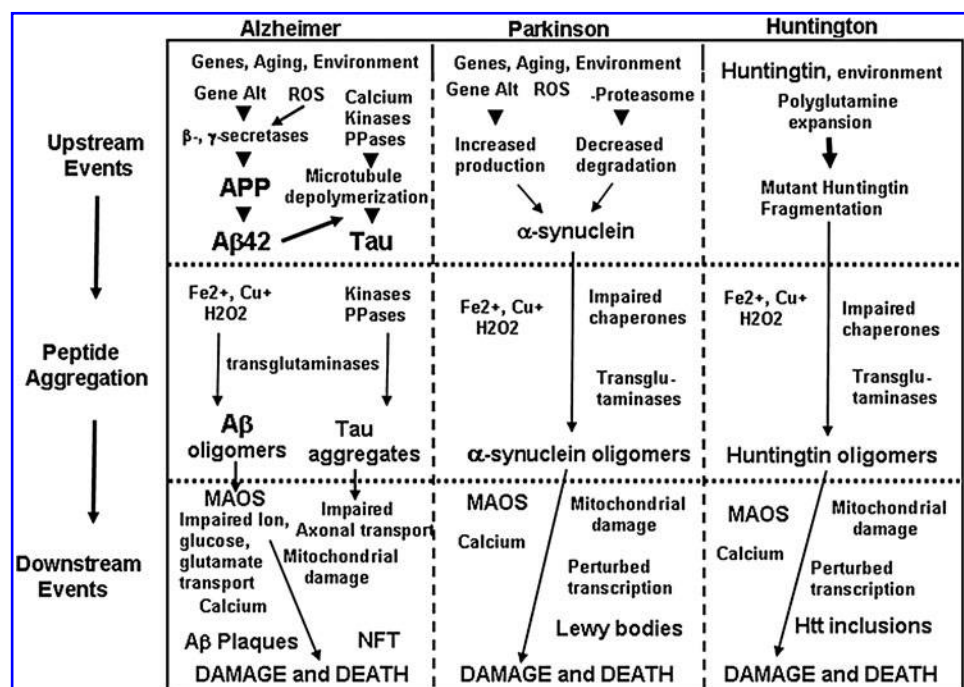
### Protection of Neurons Against Oxidative Stress by Three Behaviors that Reduce the Risk of AD

In this section we describe the now considerable evidence that the risk of AD can be reduced by three behaviors that activate ACSRP (cognitive stimulation, exercise, and dietary energy restriction). In the following section we then present evidence that these three behaviors exert their beneficial effects, at least in part, by protecting neurons against oxidative stress (Figs. 3 and 4).

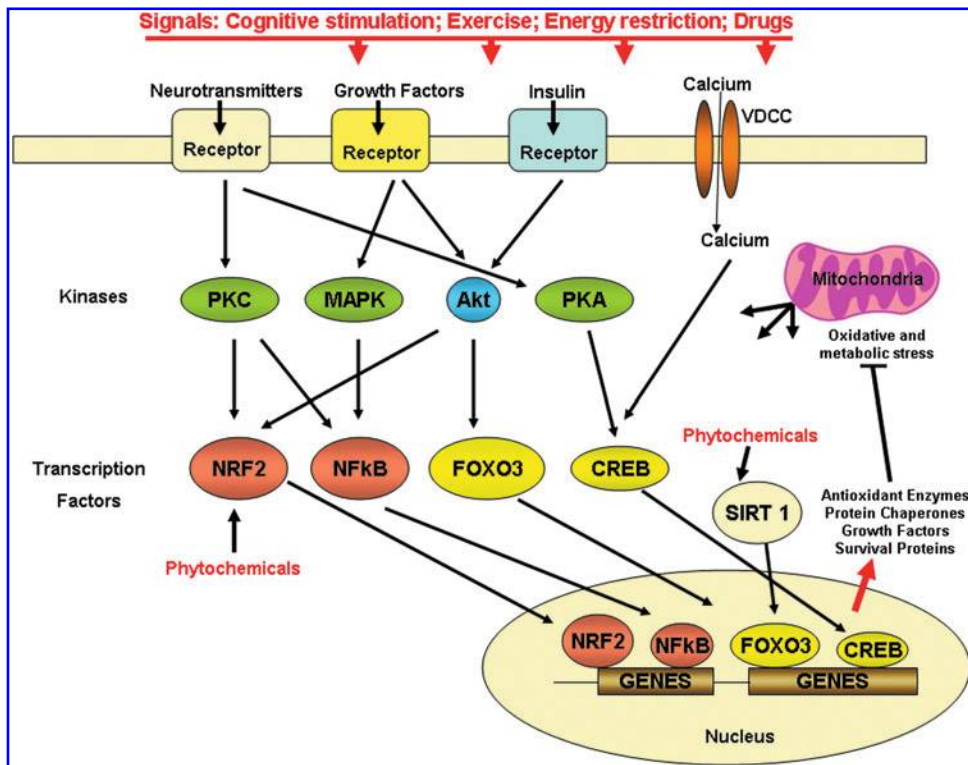
#### Human studies

In 1988 Katzman *et al.* observed that some subjects failed to develop dementia despite postmortem evidence of advanced AD pathology and suggested that this phenomenon may be due to these subjects having a greater reserve in both brain

**FIG. 2. Working model for the mechanisms of proteotoxic damage to neurons in AD, Parkinson's disease (PD), and Huntington's disease (HD).** Oxidative stress resulting from the aging process, combined with environmental and genetic factors, promotes disease-specific molecular perturbations that play key roles in the neurodegenerative cascades in AD, PD, and HD. In each disorder there are one or more pathogenic, self-aggregating proteins involved: A $\beta$  and tau in AD,  $\alpha$ -synuclein in PD, and huntingtin in HD. Events involving oxidative stress upstream and downstream of pathogenic protein aggregations are illustrated. A $\beta$ 42, amyloid  $\beta$ -peptide 1-42; Htt, huntingtin; MAOS, membrane-associated oxidative stress; NFT, neurofibrillary tangles; PPases, protein phosphatases; ROS, reactive oxygen species. Modified from Mattson and Magnus (117).

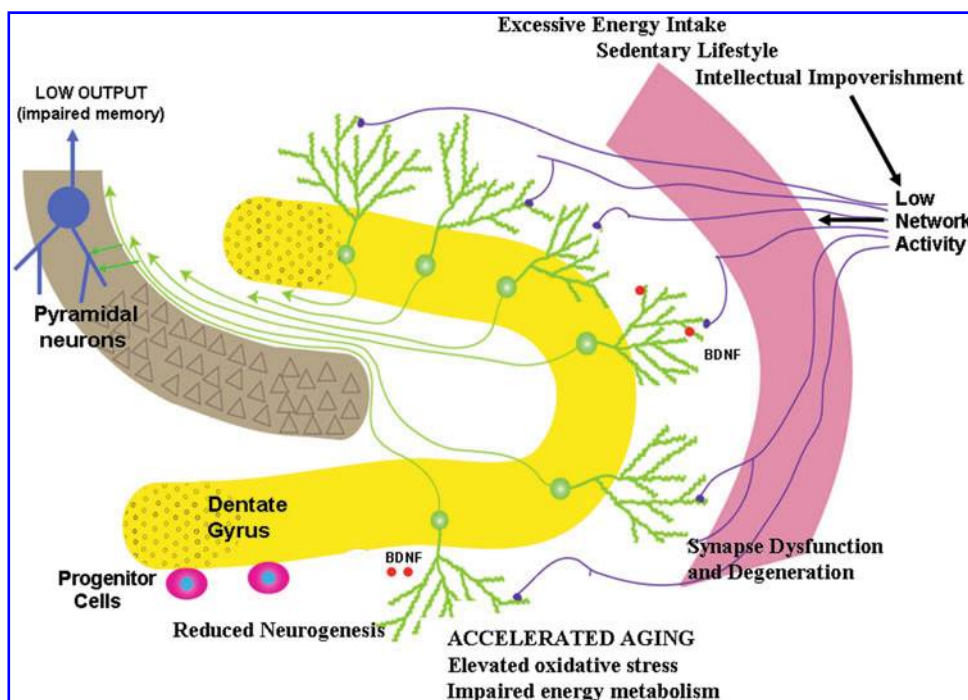






**FIG. 3. Adaptive stress response pathways that may be compromised in AD.** The increased synaptic activity in nerve cell networks involved in cognition engages several signal transduction pathways that ultimately lead to the production of proteins that protect neurons against oxidative and metabolic stress. Mental and physical exercise and dietary energy restriction are three examples of AD risk-reducing behaviors that activate neurotransmitter (e.g., glutamate, serotonin, and norepinephrine), growth factor (e.g., brain-derived neurotrophic factor [BDNF], nerve growth factor, glial cell line-derived neurotrophic factor, and vascular endothelial growth factor), and hormone (e.g., insulin, GLP-1, and ghrelin) receptors. Cognitive stimulation, exercise, and dietary energy restriction all increase activity in neuronal circuits, resulting in the activation of neurotransmitter (particularly glutamate) receptors, calcium influx, and production of growth factors. Specific kinases and transcription factors are activated that mediate adaptive cellular stress responses. The transcription factors induce expression of genes encoding, for example, antioxidant enzymes, protein chaperones, neurotrophic factors, and cell survival proteins. Certain phytochemicals may stimulate one or more adaptive cellular stress response pathways, either directly by interacting with kinases or transcription factors, or indirectly by inducing oxidative and/or metabolic stress. Modified from Mattson and Cheng (114). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

vation of neurotransmitter (particularly glutamate) receptors, calcium influx, and production of growth factors. Specific kinases and transcription factors are activated that mediate adaptive cellular stress responses. The transcription factors induce expression of genes encoding, for example, antioxidant enzymes, protein chaperones, neurotrophic factors, and cell survival proteins. Certain phytochemicals may stimulate one or more adaptive cellular stress response pathways, either directly by interacting with kinases or transcription factors, or indirectly by inducing oxidative and/or metabolic stress. Modified from Mattson and Cheng (114). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).



**FIG. 4. Adverse consequences of lifestyles that disengage adaptive cellular stress responses in the hippocampus.** A continuous positive energy balance resulting from overeating and physical underactivity, and a cognitively impoverished lifestyle all result in relatively low levels of integrated input to the hippocampus. As a result of suboptimal levels of activation of adaptive neuronal stress response pathways, network activity, there is reduced production of neurotrophic factors such as BDNF and fibroblast growth factor 2, protein chaperones, and antioxidants. Neurons are thereby rendered vulnerable to aging and metabolic stress resulting in oxidative damage, reduced synaptic plasticity, synapse loss, and impaired neurogenesis. Modified from Lazarov *et al.*, 2010 (94). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

synaptic plasticity, synapse loss, and impaired neurogenesis. Modified from Lazarov *et al.*, 2010 (94). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article at [www.liebertonline.com/ars](http://www.liebertonline.com/ars)).

mass and neuronal number (84). The idea of brain or cognitive reserve has since expanded to refer to the ability of as yet unknown genetic factors and/or increased brain use during early and midlife to afford neuroprotection in the face of aging and neurodegenerative disease. At the cellular level there is evidence for greater numbers of neurons and synapses in individuals deemed to have a high cognitive reserve (53, 127, 163). Various epidemiological studies have shown that individuals with higher levels of education can tolerate more AD brain pathology without the corresponding levels of dementia seen in individuals with less education (12, 13, 52, 136). Studies have also shown a reduced risk of dementia in patients with high job complexity or with a cognitively active lifestyle (164). Increased mental activity has been shown to be correlated with a reduced incidence of AD (174) and reduced age-associated hippocampal atrophy (164). Also, mental training has been shown to positively influence cognition longitudinally (162). Some studies even suggest that having an active social life can be protective against AD (52) and that the risk of AD doubles in lonely individuals (173). Taken together, the data suggest that cognitive reserve can be increased, but that to do so requires considerable effort, at least for those not inclined to exercise or challenge their mind.

In recent years it has become evident that like mental exercise, physical exercise is important for healthy brain function. There have been a variety of studies in humans that have shown that physical activity can benefit brain health (137). Fitness training has been shown to increase performance on cognitive tasks in older individuals (33). Exercise has also been associated with reduced cognitive impairment over a 2-year span in elderly individuals (47). Regular exercise is associated with a delay in the onset of dementia and AD (93, 140) and a reduced risk of developing AD (99). In particular, it was found that AD patients tend to be less active during midlife than healthy controls, suggesting that low activity level in midlife could be a risk factor for AD (54). While exercise intensities vary among individuals and from study to study, it was reported that a regular walking routine in elderly men was associated with a reduced risk of dementia (1). Colcombe *et al.* showed that both gray and white matter increased in healthy older adults as a function of fitness level, but the same correlation was not seen in younger subjects (34). Also, cardiorespiratory fitness was shown to be positively correlated with medial temporal cortex volume in AD patients, but not in healthy controls, suggesting that cardiorespiratory fitness may modify AD-related brain atrophy (75). Most of the studies on fitness and cognition look at leisure time activities, but one study showed that work-related physical activity alone was not protective against dementia or AD later in life (141).

In addition to keeping physically active there is evidence that maintaining a healthy body weight can also benefit cognition. Studies have shown that increased body mass index (BMI) is associated with a decrease in cognitive performance in healthy individuals (63) and that long-term adult obesity is associated with lower cognition scores (143). Gunstad *et al.* observed that whole brain and gray matter volumes were reduced in obese individuals compared with normal or moderately overweight controls (62). One complication of relating BMI or body fat levels to cognition is that many overweight subjects also have diabetes, which has been shown to adversely affect cognition. Several studies have

shown that the risk of dementia and AD is increased in individuals with diabetes (5, 20, 131). Even impairment in glucose regulation that leads to borderline diabetes has been shown to increase the risk of dementia and AD regardless of the future progression to diabetes proper (177). The most effective intervention for both diabetes and obesity is dietary energy restriction. It was reported that individuals with relatively low caloric intakes are at reduced risk for AD (104) and adherence to a Mediterranean diet has been associated with a lower risk for mild cognitive impairment and AD (144).

### Animal studies

The usual housing conditions for laboratory mice and rats result in animals with relatively little cognitive reserve as they age; the animals are overfed, sedentary, and cognitively unchallenged (105). This is readily apparent by the results of studies in which the animals are fed less, exercise, and/or are maintained in enriched environments. Environmental enrichment (EE) paradigms vary, but usually involve housing the animals in large cages with a variety of objects to explore and increased opportunities for social interactions (166). In the late 1940s Hebb documented behavioral differences between rats kept at his home compared with rats kept in the laboratory, and since then a number of studies have established behavioral, anatomical, and molecular changes in rodents following EE (166). The changes seen include improved performance in learning and memory tasks, increased brain mass, enhanced neurogenesis, and changes in dendrite numbers (166). Studies looking at older dogs have shown that when fed a diet high in antioxidants and exposed to EE, the dogs exhibit enhanced cognitive performance, less oxidative protein damage, and high endogenous antioxidant activity than either treatment alone (35a). Enrichment alone was able to improve cognition in older dogs compared with non-enriched controls.

EE has also been studied in transgenic models of AD (6, 14, 35, 40, 80). Several investigators have reported reductions in A $\beta$  levels (6) and amyloid deposits (95) in transgenic AD mouse models after EE, whereas one study reported an increase in amyloid burden (81). Learning and memory was improved after EE in single-mutant APPswe and PS1 mice, whereas double-mutant APPswe/PS1 mice had a more limited improvement (81). Both cognitive function improvements and decreases in neuropathology have been seen in mice that are exposed to an enriched environment at a young age; this is seen in PS1/DAPP mice (35), AD11 mice (14), and PS1 and PS2 KO mice (40). APPswe mice placed in EE at a later age (16 months) showed improvements in cognitive function compared with home cage controls, but without a decrease in A $\beta$  levels (10). Similar findings were reported for APP23 mice that started EE at 10 weeks of age and showed improved water maze performance but stable A $\beta$  levels (175). These results suggest at least two different effects of EE on AD pathogenesis, a reduction A $\beta$  accumulation when EE is initiated early in life, and improvement of cognitive function when EE is started before or after A $\beta$  pathology has already developed.

EE has also been shown to improve neurogenesis in AD mouse models and boost hippocampal long-term potentiation (LTP) (76). Herring *et al.* found that after cognitive stimulation there was an increase in the number of newborn and mature hippocampal neurons and in molecules associated with

plasticity in a mouse model of AD. The levels of neurogenesis in AD mice raised in an enriched environment were comparable to those of nontransgenic control mice (71). Changes in oxidative stress have also been reported after EE, with decreases in ROS and markers of oxidative damage, and increases in antioxidant defense mechanisms (72).

In an alternative form of cognitive stimulation, Billings *et al.* used serial trainings of 3xTgAD mice in the water maze mice and obtained improvements similar to those achieved with EE. Improved memory performance was seen in these mice compared with control animals that were allowed to swim without learning a platform location, but like the EE studies training had to be initiated early, before overt pathology developed (21).

One of the complications of EE is that many paradigms involve the addition of running wheels. Some studies have looked at EE compared with running alone and saw no enhancement of spatial learning or neurogenesis with running alone (175). Others have looked at different combinations of social, physical, and cognitive stimulation and found that physical and social interaction were alone not enough to protect against cognitive decline in APP mutant transgenic mice (36). On the contrary, it has been shown in young C57BL/6 females that neurogenesis, as measured by BrdU incorporation, induced by running alone was equivalent to that in mice exposed to EE with access to a running wheel (167).

Comparing exercise and enrichment in young C57BL/6 female mice, Harburger *et al.* showed that only exercise improved spatial memory in young mice, whereas both exercise and enrichment improved memory in middle-aged or old mice (68). Other studies have shown an increase in neurogenesis after 3 but not 1 h of running (74). The actual number of new cells born was positively correlated with the distance run among mice (135), but there were no significant correlations between the amount of running and a variety of behaviors, including emotionality, exploratory activity, sensory-motor processing, and spatial memory, suggesting that, at least within the limits of this study, running does not have major effects on the behavior of young mice (132). In young C57BL/6 mice, 7–10 days' running was enough to enhance LTP expression (169). Long-term running for 94 weeks in male rats reduced oxidation levels of DNA and lipids in the cerebellum; reduction of lipid oxidation was seen as early as 3 months of running and correlated with forelimb grip strength (37). Radak *et al.* observed similar findings, increased memory, decreased protein carbonyls, and increased proteasome complex activity in the brains of middle-age rats that were allowed to swim for an hour a day, 5 days a week for 9 weeks (134).

Exercise has also been shown to be beneficial to mouse models of AD (3, 124–126). In old Tg2576 mice, access to a running wheel for 13 weeks improved memory to levels similar to nontransgenic animals (125). Other studies found that as little as 3 weeks' running in 15–19-month-old Tg2576 mice, a time point where significant AD pathology has begun, was able to improve spatial learning (129) and decrease level soluble forms of A beta (126). One month of running in young TgCRND4 mice resulted in decreased proteolytic fragments of APP and after 5 months decreased A $\beta$  plaques in cortex and hippocampus (3). The benefits seen with exercise appear to be greater with voluntary exercise compared with forced exercise. While both forced and voluntary running in Tg2576 mice result in larger hippocampal volumes, only voluntary runners

show an increase in memory function and a decreased in A $\beta$  plaques (183).

In addition to physical and mental activity, body weight and energy intake have been shown to be correlated with health and life-span (105, 106). Young rats on a high-caloric diet for 6 weeks gained weight and exhibited learning and memory deficits (82). Diets with high saturated fat content impaired cognitive function in a delayed alternation task in young rats, and the percent of saturated fat in the diet correlated positively with behavioral impairment (61). Murray *et al.* showed that only 9 days of a high-fat diet was sufficient to cause physical impairment on a treadmill and cognitive impairment in the water maze (121). When 16-month-old rats were fed diets high in fat and cholesterol they made more errors in a test of working memory especially when memory loads were high, and they also showed altered hippocampal morphology (60). Similar studies of mice showed that high-fat diets worsen performance in learning and memory tests (49, 181). There is some evidence that the effects of obesity on neuroplasticity and cognitive function differ in males and females. After 4 weeks on a high-fat diet, hippocampal neurogenesis was impaired in male but not female rats (98). Obese male mice, but not obese female mice, showed memory deficits and impaired level of LTP and long-term depression (77). Evidence is emerging that the weight of the pregnant dam can increase dentate gyrus lipid peroxidation and impair neurogenesis in her offspring (160). Rats that were kept on a high-fat diet after being born to dams on high-fat diets had increased susceptibility to memory impairment and increase oxidative stress compared with rats that were either kept on a high-fat diet after being birthed by a dam on a normal diet, or rats kept on a normal fat diet after being birthed by a dam on a high fat (171).

The impact of energy intake on the brain is further appreciated when one considers the results of studies in which animals are maintained on reduced energy diets, affected either by limited daily feeding/caloric restriction (CR) or intermittent fasting (IF)/alternate day fasting (106). When the daily energy intake of mice was reduced by 20%, their performance on a learning and memory task was improved (69). IF for 6–8 months in mice resulted in enhanced learning and synaptic efficiency, which was associated with increased expression of N-methyl-D-aspartate receptors in the hippocampus and perirhinal cortex (51). CR in rats prevents age-related cognitive decline in old but not young rats, and maintains levels of N-methyl-D-aspartate and alpha-amino-5-methyl-3-hydroxy-4-isoxazolepropionic acid receptors in the hippocampus, which otherwise decrease during aging (2, 148). On the other hand, CR lasting 7–24 months in rats was reported to increase longevity but had a negative impact on cognition (180). In the 3xTgAD mouse model of AD, IF and CR beginning at 5 months of age and lasting for 1 year improved water maze performance and exploratory behavior compared with mice fed AL (67). Mice on CR had lower levels of A $\beta$ 1-40, A $\beta$ 1-42, and phospho-tau in the hippocampus compared with mice on AL or IF diets (67).

One common complication of excess energy intake is diabetes, which itself has been shown to affect cognition adversely. Diabetic rats have been shown to have impaired spatial learning and LTP (18, 19). Treatment of the diabetic rats with insulin after cognitive decline was not able to ameliorate the deficits (19). Insulin-resistant rats also demonstrate reduced spine density in the hippocampus, reduced LTP,



impaired spatial learning, and a reduction in hippocampal neurogenesis (151, 154). In the APP/PS1 mouse model of AD, insulin resistance can be induced by adding 10% sucrose to the drinking water, mimicking type II diabetes. Compared with control transgenic mice fed normal water the insulin-resistant group developed greater memory impairment and increases in A $\beta$  deposition (26). In the Tg2576 mouse model of AD, diet-induced insulin resistance resulted in increase A $\beta$  plaque formation and a decrease insulin receptor signaling (73).

### Molecular Mechanisms of Adaptive Responses to Oxidative Stress

We believe and have proposed previously that cognitive stimulation, exercise, and dietary energy restriction promote neuronal survival and plasticity by activating adaptive cellular stress responses in neurons (105, 110). When neurons are engaged in cognitive processes or in controlling body movements, their electrical and synaptic activity results in Na<sup>+</sup> and Ca<sup>2+</sup> influx, and an increased energy demand. This ionic and energetic stress also results in increased production of ROS, including mitochondrial and extramitochondrial superoxide, hydrogen peroxide, and nitric oxide (115). Similar events occur in many neurons during exercise and when dietary energy intake is low. Normally, neurons respond to this mild stress adaptively, as indicated at the molecular level by their upregulation of expression of genes encoding neurotrophic factors such as BDNF and FGF2, protein chaperones such as HSP70 and GRP78, and antioxidant enzymes such as HO-1 (11). The mechanisms by which these cellular defenses are mobilized are beginning to be understood and are described below (Fig. 3).

Several signal transduction pathways have been implicated in the mechanisms by which neurons respond adaptively to mental and physical exercise, and dietary energy restriction. The transcription factors CREB, NF- $\kappa$ B, and Nrf2 are activated in response to vigorous synaptic activity, and by energetic and oxidative stress (100, 149, 168). CREB is activated by Ca<sup>2+</sup> and then induces expression of several neuroprotective proteins, among which BDNF has been the most heavily studied with regards to roles in the adaptive responses of neurons to cognitive stimulation, exercise, and energy restriction (118).

#### *Neurotrophic factors and the battle against oxidative stress: BDNF as a prototype*

BDNF plays pivotal roles in synaptic plasticity and neurogenesis, and can protect neurons against excitotoxic, oxidative, and metabolic stress (32, 150). BDNF has been shown to increase the production of the antioxidant enzyme glutathione peroxidase 1 (116) and the membrane-associated antioxidant protein Bcl-2 (23, 145). BDNF was originally identified as a neurotrophin that plays key roles in development of the nervous system, and has since been shown that BDNF and the high affinity BDNF receptor trkB are widely expressed in neurons throughout the brain and spinal cord (158). BDNF levels have been shown to be increase upon LTP induction (28) and BDNF promotes changes at dendritic spine structure (157). Suppression of BDNF production can block dendritic structural changes (157), and mice lacking BDNF in their forebrain neurons exhibit impaired LTP (89, 90) and learning and memory (59). Administration of exogenous

BDNF to knockout hippocampal slices abrogated LTP impairment (130). Thus, there is strong evidence to implicate BDNF as an important mediator of synaptic plasticity.

BDNF has also been implicated to play a role in the age-related changes in brain morphology and memory decline. In some human studies BDNF levels in serum (188) or plasma (103) exhibited a negative correlation with age. An analysis of plasma BDNF levels in 496 middle-aged and elderly subjects from the Baltimore Longitudinal Study of Aging demonstrated a negative correlation between plasma BDNF levels and age in both males and females (56). The latter study further demonstrated that plasma BDNF levels are positively associated with risk factors for metabolic syndrome and cardiovascular disease, independently of age. Studies of the brain have shown that levels of BDNF (133) and its receptor TrkB (138) decrease over the lifespan. Although there have been some inconsistent results, most studies in animals have demonstrated similar decreases in BDNF and TrkB with age, and have also shown that decreased brain BDNF levels are correlated with impaired memory and decreased dendritic spine density in hippocampal neurons (157). Thus, reduced BDNF levels in the brain render neurons vulnerable to dysfunction and degeneration, whereas the functions of BDNF in the blood are unknown. In AD, BDNF and TrkB levels are reduced in several areas of the brain, even at preclinical stages of AD (157), although at least one study reported an increase in BDNF levels in the hippocampus of AD patients (45). BDNF levels in the APP23 transgenic mouse model of AD are dissimilar to human studies, whereas hippocampal BDNF levels are lower in APP23 mice, levels in the frontal cortex are elevated (70), and in cortex and striatum increases were age-dependent (146). Because neuronal death does not occur in the latter mouse models of AD, it is possible that the increased BDNF levels may protect the neurons against A $\beta$ ; BDNF can indeed protect neurons against A $\beta$  toxicity in experimental models (9).

In conjunction with increased memory performance and neurogenesis, EE has been shown to increase in hippocampal BDNF levels in both mice (186) and rats (79). In BDNF heterozygote mice with reduced levels of BDNF, the effectiveness of EE in inducing neurogenesis is reduced (139), and the BDNF<sup>+/-</sup> mice also exhibit reduced dendritic spine density in CA1 and dentate gyrus neurons compared with wild-type mice (185). In the APP23 mouse model of AD, improvements in water maze performance and hippocampal neurogenesis from EE are accompanied by increased hippocampal expression of BDNF (175). The data described above confirm the necessity of BDNF for the synaptic plasticity and neurogenesis induced by EE.

Exercise is beneficial for brain health in humans (165). In one study serum BDNF levels were elevated in response to exercise and the BDNF levels were positively correlated with cognitive performance (50). Voluntary running has also been shown to increase levels of BDNF in the hippocampus of rats (122). Increased BDNF levels in rats after exercise were also associated with increases in memory performance and increases in proteins involved in energy metabolism such as AMP-activated protein kinase, insulin-like growth factor 1, and ghrelin (57). These increases were prevented when BDNF was blocked during exercise. In mice there were positive correlations between BDNF, CREB, and learning rates after exercise (170). In the same study it was found that inhibition of

BDNF during running prevents the enhanced memory function associated with running. Other studies in mice have shown that BDNF levels are increased in the hippocampus within 3 weeks of voluntary exercise and levels remain high up to 2 weeks after the end of the exercise period (15). BDNF levels did not return to baseline until after 3–4 weeks after the running period, and the performance in the radial arm maze was best when performed 1 week after the end of running. The SynRas transgenic mouse with permanently activated Ras is expressed under the neuronal synapsin I promoter and has been shown to have decreased neurogenesis and impaired short-term memory (92). When these mice were allowed to run, they showed increased basal BDNF levels comparable to running wild-type animals and also an amelioration of both neurogenesis and short-term memory deficits (92). In the same study the authors looked at TrkB and doublecortin costaining and found that TrkB staining occurred in immature proliferative cells and those with complex dendritic arbors, suggesting that BDNF can act on newly born neurons at multiple stages (92). In diabetic db/db mice, increased BDNF levels from running were accompanied by increase dendritic spine density in dentate granule neurons (153). In the NSE/APPsw mouse model of AD, after 16 weeks of voluntary running, increased brain BDNF levels were complemented by a decrease in A $\beta$ 42 peptide and decreased markers of apoptosis (161).

Emerging evidence has suggested that BDNF may regulate energy balance. Human studies have shown that serum BDNF levels are increased in obese women and decreased in women with anorexia nervosa compared with healthy individuals (119). Epidemiological evidence suggests that polymorphisms in BDNF may affect weight; the Val66Met BDNF polymorphism is associated with a lower BMI in healthy subject compared with healthy individuals with alternate polymorphisms (64). Mice with reduced BDNF levels (BDNF<sup>+/-</sup> mice) exhibit increased food intake, insulin resistance/diabetes, and obesity (43, 87). The metabolic abnormalities of BDNF<sup>+/-</sup> mice can be reversed with intraventricular infusion of BDNF in the brain and by dietary energy restriction (43, 87). BDNF<sup>+/-</sup> mice also display impaired hippocampal neurogenesis, which can be partially restored by dietary energy restriction (97). Diets high in saturated fat have been shown to decrease BDNF levels and impair cognitive function, and administration of the antioxidant Vitamin E in conjunction with the high-fat diet was able to restore levels of BDNF and CREB activity, and also reversed cognitive impairment, suggesting that oxidative stress mediates the adverse effects of a high-fat diet on BDNF signaling and cognitive function (176).

BDNF has also been shown to help regulate glucose metabolism in mouse models of diabetes. Subcutaneous administration of BDNF in db/db mice was able to decrease blood glucose and body weight; these changes remained for weeks after BDNF treatment, suggesting the induction of physiological changes (159). Subcutaneous administration of BDNF when started early (4 weeks) was able to prevent the age-related increases in blood glucose levels seen in db/db mice (179). Others have shown that intracerebroventricular administration of BDNF can lower glucose levels and increase insulin concentrations in the pancreas in db/db mice, suggesting the CNS had a role in glucose metabolism (128). CR in db/db mice increases hippocampal BDNF levels and is asso-

ciated with increased dendritic spine density (153). Taken together, the data suggest that the cognitive impairment observed in diabetes may be a result of dysregulation of glucose metabolism due to decreased BDNF levels.

### Protein Chaperones, Antioxidants, and Adaptive Cellular Stress Responses

As their name implies, a major function of protein chaperones is to bind to other proteins and protect them from being exposed to adverse factors, including oxidative stress. The prototypical protein chaperone is HSP70, which has been shown to be upregulated in neurons in response to a range of insults, including cerebral ischemia and epileptic seizures (22). Failure of protein chaperone-mediated neuroprotection is implicated in the pathogenesis of several neurodegenerative disorders, including Parkinson's disease, AD, and Huntington's disease (Fig. 2). Both dietary energy restriction and 2-deoxyglucose administration were reported to upregulate the expression of GRP78 and HSP70 and protect dopaminergic neurons against the toxicity of chemical inhibitors of mitochondrial complex I (44). Multiple protein chaperones, including HSPs 70, 40, and 90, can protect neurons against polyglutamine-induced degeneration in models relevant to Huntington's disease (55).

It was reported that the protein co-chaperone BAG2 forms a complex with HSP70, which then binds insoluble/hyperphosphorylated tau and delivers it to the proteasome for degradation (27). Others have demonstrated a role for HSP90 in the degradation of phosphorylated tau (38). HSP70 and HSP90 can inhibit the aggregation of A $\beta$ 1-42 *in vitro* (48), suggesting a potential role for these protein chaperones in protecting brain cells against AD by preventing the aggregation and neurotoxicity of A $\beta$ . It was reported that addition of HSP90, HSP70, and HSP32 to the culture medium induces the production of interleukin 6 and tumor necrosis factor, and increases the phagocytosis and clearance of A $\beta$ , by microglia (83). Impaired chaperone function and consequent proteotoxicity may play a role in the selective vulnerability of neurons in different brain regions in AD. As evidence, the levels of immunostaining for HSP72 and proteasomal subunits are weaker in neurons in brain regions affected in AD (4).

Not only does expression of HSPs increase in muscle after exercise (58, 120), but increased expression also occurs in neurons in response to exercise (31) and dietary energy restriction (11, 182). In an animal model of heat stroke-induced hyperthermia, 3 weeks of exercise induced HSP72 expression in the brain and protected neurons against damage and also increased the survival of the animals (31). Alternate day fasting for several weeks to months results in the upregulation of HSP70 and HSP40 in hippocampal pyramidal neurons in mice and rats, and protects those neurons against excitotoxic death in experimental models of severe epileptic seizures (24, 147). A moderate level of pharmacologically induced energetic stress has also been shown to upregulate protein chaperones and protect neurons against oxidative and excitotoxic injury. For example, treatment of hippocampal neurons with 2-deoxyglucose, a form of glucose that inhibits glycolysis, results in increased expression of HSP70 and GRP78, and protects the neurons from being killed by oxidative (Fe<sup>2+</sup>) and excitotoxic (glutamate) insults (96). Similarly, exposure of hippocampal neurons to iodoacetate, an inhibitor of glycer-



aldehyde-3-phosphate dehydrogenase, induces expression of HSP70, HSP90, and the antioxidant protein Bcl-2, and protects the neurons against oxidative injury (66).

### Mitochondrial Neurohormesis

Mitochondria produce a major portion of free radicals generated in nerve cells; during oxidative phosphorylation, superoxide anion radical is produced. SOD within mitochondria (SOD2) and in the cytosol (SOD1) convert superoxide to hydrogen peroxide (111). Hydrogen peroxide can be converted to water by the activities of catalase and glutathione peroxidase; however, in the presence of  $\text{Fe}^{2+}$  or  $\text{Cu}^{+}$ , the hydrogen peroxide is converted to hydroxyl radical. Hydroxyl radical can be very damaging to proteins and nucleic acids, and to membranes in which it attacks double bonds in fatty acids to initiate a chain reaction called lipid peroxidation. Another chemical pathway that induces oxidative damage involves  $\text{Ca}^{2+}$ -induced activation of nitric oxide synthase resulting in the production of nitric oxide, a free radical. Nitric oxide can, in turn, interact with superoxide to generate peroxynitrite, which damages proteins by causing the nitration of tyrosine residues; peroxynitrite can also induce membrane lipid peroxidation. Increased levels of superoxide, hydrogen peroxide, hydroxyl radical, nitric oxide, and peroxynitrite have been suggested to contribute to the excessive oxidative damage to proteins, nucleic acids, and membranes documented in studies of postmortem tissue from AD patients, and in experimental models of AD (142, 155). SOD2 can protect neurons against insults relevant to AD, including  $\text{A}\beta$ ,  $\text{Fe}^{2+}$ , and nitric oxide-generating agents (86).

In addition to SOD2, mitochondria contain several proteins that help protect neurons against oxidative damage. One class of such proteins are the mitochondrial uncoupling proteins (UCPs), which are integral membrane proteins in the mitochondrial inner membrane that provide a conduit for leakage of protons across the membrane thereby reducing oxidative phosphorylation and the production of superoxide. Recent studies have shown that overexpression of UCP4 (101) and UCP2 (109) can protect neurons against oxidative insults relevant to AD, including glycolytic inhibitors and mitochondrial toxins. UCP4 was also shown to stabilize cellular and mitochondrial calcium homeostasis, which was associated with reduced levels of mitochondrial ROS and increased resistance of neural cells to endoplasmic reticulum stress (30). Neuronal UCPs can be activated by free fatty acids and oxidative stress, and UCPs may regulate cellular calcium homeostasis, free radical production and mitochondrial biogenesis (8). The latter article reviews additional findings further suggest roles for UCPs in synaptic plasticity and neurodegenerative disorders.

Environmental factors that protect the brain against cognitive impairment and AD have been shown to increase expression SOD2 and UCPs. For example, both exercise and CR have shown to increase levels of UCPs in the brain (39, 101).

Experimental reduction in SOD2 levels in a transgenic mouse model of AD resulted in an acceleration of the development of  $\text{A}\beta$  pathology and cognitive deficits (46). The function of SOD2 may be impaired in AD because it was shown that  $\text{A}\beta$  can induce the nitration of SOD2 in APP/PS1 double-mutant AD mouse model (7). When mice with AD-like pathology were maintained in an enriched environment,

the amount of oxidative stress associated with the  $\text{A}\beta$  pathology was significantly reduced, a result that was apparently due to the upregulation of antioxidant defenses, including SOD1 and SOD2, in the brain cells (72).

In our study called atlas of gene expression in mouse aging project, male and female mice that had been maintained on either an *ad libitum* diet or a 40% CR diet beginning at 6 weeks of age were killed at 6, 16, and 24 months of age, and expression of nearly 17,000 genes was determined in 16 different tissues (178, 184). Our analyses of the CNS revealed that aging is associated with downregulation of genes encoding proteins involved in DNA repair, protein degradation, and inhibitory neurotransmission, and that CR counteracts these effects of aging (178). In another study we performed a gene array analysis of the hippocampus in male and female rats that had been maintained for 6 months on either *ad libitum* (control), 20% CR, 40% CR, IF, or high fat/high glucose diets. The CR diets significantly increased the size of the hippocampus of females, but not males, and this gender difference was associated with specific changes in hippocampal gene expression (107). The 20% CR diet downregulated genes involved in mitochondrial energy production in males, while upregulating these metabolic pathways in females. The 40% CR diet upregulated genes involved in glycolysis, protein deacetylation, PGC-1 $\alpha$ , and mTor pathways in both sexes. Genes involved in energy metabolism, oxidative stress responses, and cell survival were affected by the high-energy diet in both males and females. Collectively, these findings suggest that aging results in dysregulation of mitochondrial energy and oxyradical metabolism resulting in reduced energy availability and increased ROS production and oxidative damage.

Exercise may protect the brain against AD by stimulating ACSRP. To elucidate the mechanisms by which exercise may benefit the brain during aging, we trained 16-month-old mice that had either run regularly during their adult life or led a sedentary lifestyle in the hippocampus-dependent water maze (152). We then analyzed expression of 24,000 genes in the hippocampus and found that runners show greater activation of genes associated with synaptic plasticity and mitochondrial function, and also exhibit significant downregulation of genes associated with oxidative stress and lipid metabolism. These results suggest that the enhancement of cognitive function by lifelong exercise is associated with preservation of mitochondrial function and suppression of oxidative stress.

### Therapeutic Implications

While research on neuronal plasticity, aging, and AD is ever-expanding, a pharmacological cure for dementia is not imminent. The present mini-review of the current literature does, however, offer approaches that could be beneficial in preventing or delaying the onset of dementia. In line with the hormesis ideology—"what doesn't kill you makes you stronger"—upregulation of adaptive stress responses in cells *via* cognitive stimulation, exercise, and dietary restriction better prepare the brain for oxidative insults resulting from aging and disease (93, 110, 140, 164). On the other hand, lifestyles that include social isolation, low activity levels, and obesity may hasten the onset of cognitive impairment and AD (54, 63, 173). Upregulation of specific proteins that enhance neurogenesis and promote cell survival may be key

modulators of the adaptive stress response. Data suggest that these molecular pathways may be impaired in certain disease states, including AD.

Activation of adaptive stress responses in animal models of neurodegenerative diseases have been able to provide benefits in cognition and motor function, and suggest that these molecular pathways may be able to be activated through cognitive stimulation, exercise, and CR (41, 42, 67, 76, 108, 125). One of the key factors that may help protect against the decline of cognitive function in aging and disease is BDNF. Experiments have shown that increasing levels of BDNF either by exogenous administration or *via* cognitive stimulation, exercise, and CR can help enhance neurogenesis and promote cognitive function (9, 158, 179). Moreover, it may be possible to induce BDNF production with pharmacological agents as exemplified by the most widely used and efficacious antidepressants such as fluoxetine, sertraline, and paroxetine, which upregulate BDNF expression and preserve neuronal function in animal models of AD (123) and Huntington's disease (41, 44a, 61a). Further research will expand our understanding of how adaptive stress responses can modulate synaptic plasticity and improve cognitive decline with aging and disease, and how to tap into these pathways to suppress oxidative damage and protect the brain against AD.

### Acknowledgment

This work was supported by the Intramural Research Program of the National Institute on Aging, NIH.

### References

- Abbott RD, White LR, Ross GW, Masaki KH, Curb JD, and Petrovitch H. Walking and dementia in physically capable elderly men. *JAMA* 292: 1447–1453, 2004.
- Adams MM, Shi L, Linville MC, *et al.* Caloric restriction and age affect synaptic proteins in hippocampal CA3 and spatial learning ability. *Exp Neurol* 211: 141–149, 2008.
- Adlard PA, Perreau VM, Pop V, and Cotman CW. Voluntary exercise decreases amyloid load in a transgenic model of Alzheimer's disease. *J Neurosci* 25: 4217–4221, 2005.
- Adori C, Kovács GG, Low P, Molnár K, Gorbea C, Fellingner E, Budka H, Mayer RJ, and László L. The ubiquitin-proteasome system in Creutzfeldt-Jakob and Alzheimer disease: intracellular redistribution of components correlates with neuronal vulnerability. *Neurobiol Dis* 19: 427–435, 2005.
- Akomolafe A, Beiser A, Meigs JB, *et al.* Diabetes mellitus and risk of developing Alzheimer disease: results from the Framingham study. *Arch Neurol* 63: 1551–1555, 2006.
- Ambree O, Leimer U, Herring A, *et al.* Reduction of amyloid antipathy and abeta plaque burden after enriched housing in TgCRND8 mice: involvement of multiple pathways. *Am J Pathol* 169: 544–552, 2006.
- Anantharaman M, Tangpong J, Keller JN, Murphy MP, Markesbery WR, Kinningham KK, and St. Clair DK. Beta-amyloid mediated nitration of manganese superoxide dismutase: implication for oxidative stress in a APPNLH/NLH X PS-1P264L/P264L double knock-in mouse model of Alzheimer's disease. *Am J Pathol* 168: 1608–1618, 2006.
- Andrews ZB, Diano S, and Horvath TL. Mitochondrial uncoupling proteins in the CNS: in support of function and survival. *Nat Rev Neurosci* 6: 829–840, 2005.
- Arancibia S, Silhol M, Mouliere F, *et al.* Protective effect of BDNF against beta-amyloid induced neurotoxicity *in vitro* and *in vivo* in rats. *Neurobiol Dis* 31: 316–326, 2008.
- Arendash GW, Garcia MF, Costa DA, Cracchiolo JR, Wefes IM, and Potter H. Environmental enrichment improves cognition in aged Alzheimer's transgenic mice despite stable beta-amyloid deposition. *Neuroreport* 15: 1751–1754, 2004.
- Arumugam TV, Phillips TM, Cheng A, Morrell CH, Mattson MP, and Wan R. Age and energy intake interact to modify cell stress pathways and stroke outcome. *Ann Neurol* 67: 41–52, 2010.
- Bennett DA, Schneider JA, Wilson RS, Bienias JL, and Arnold SE. Education modifies the association of amyloid but not tangles with cognitive function. *Neurology* 65: 953–955, 2005.
- Bennett DA, Wilson RS, Schneider JA, *et al.* Education modifies the relation of AD pathology to level of cognitive function in older persons. *Neurology* 60: 1909–1915, 2003.
- Berardi N, Braschi C, Capsoni S, Cattaneo A, and Maffei L. Environmental enrichment delays the onset of memory deficits and reduces neuropathological hallmarks in a mouse model of Alzheimer-like neurodegeneration. *J Alzheimers Dis* 11: 359–370, 2007.
- Berchtold NC, Castello N, and Cotman CW. Exercise and time-dependent benefits to learning and memory. *Neuroscience* 167: 588–597, 2010.
- Bezprozvanny I and Mattson MP. Neuronal calcium mis-handling and the pathogenesis of Alzheimer's disease. *Trends Neurosci* 31: 454–463, 2008.
- Bharadwaj PR, Dubey AK, Masters CL, Martins RN, and Macreadie IG. Abeta aggregation and possible implications in Alzheimer's disease pathogenesis. *J Cell Mol Med* 13: 412–421, 2009.
- Biessels GJ, Kamal A, Ramakers GM, *et al.* Place learning and hippocampal synaptic plasticity in streptozotocin-induced diabetic rats. *Diabetes* 45: 1259–1266, 1996.
- Biessels GJ, Kamal A, Urban IJ, Spruijt BM, Erkelens DW, and Gispen WH. Water maze learning and hippocampal synaptic plasticity in streptozotocin-diabetic rats: effects of insulin treatment. *Brain Res* 800: 125–135, 1998.
- Biessels GJ, Staekenborg S, Brunner E, Brayne C, and Scheltens P. Risk of dementia in diabetes mellitus: a systematic review. *Lancet Neurol* 5: 64–74, 2006.
- Billings LM, Green KN, McGaugh JL, and LaFerla FM. Learning decreases A beta\*56 and tau pathology and ameliorates behavioral decline in 3xTg-AD mice. *J Neurosci* 27: 751–761, 2007.
- Brown IR. Heat shock proteins and protection of the nervous system. *Ann N Y Acad Sci* 1113: 147–158, 2007.
- Bruce-Keller AJ, Begley JG, Fu W, Butterfield DA, Bredesen DE, Hutchins JB, Hensley K, and Mattson MP. Bcl-2 protects isolated plasma and mitochondrial membranes against lipid peroxidation induced by hydrogen peroxide and amyloid beta-peptide. *J Neurochem* 70: 31–39, 1998.
- Bruce-Keller AJ, Umberger G, McFall R, and Mattson MP. Food restriction reduces brain damage and improves behavioral outcome following excitotoxic and metabolic insults. *Ann Neurol* 45: 8–15, 1999.
- Butterfield DA, Reed T, Newman SF, and Sultana R. Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. *Free Radic Biol Med* 43: 658–677, 2007.

26. Cao D, Lu H, Lewis TL, and Li L. Intake of sucrose-sweetened water induces insulin resistance and exacerbates memory deficits and amyloidosis in a transgenic mouse model of Alzheimer disease. *J Biol Chem* 282: 36275–36282, 2007.
27. Carrettiero DC, Hernandez I, Neveu P, Papagiannakopoulos T, and Kosik KS. The cochaperone BAG2 sweeps paired helical filament-insoluble tau from the microtubule. *J Neurosci* 29: 2151–2161, 2009.
28. Castren E, Pitkanen M, Sirvio J, *et al.* The induction of LTP increases BDNF and NGF mRNA but decreases NT-3 mRNA in the dentate gyrus. *Neuroreport* 4: 895–898, 1993.
29. Cecarini V, Ding Q, and Keller JN. Oxidative inactivation of the proteasome in Alzheimer's disease. *Free Radic Res* 41: 673–680, 2007.
30. Chan SL, Liu D, Kyriazis GA, Bagsiyao P, Ouyang X, and Mattson MP. Mitochondrial uncoupling protein-4 regulates calcium homeostasis and sensitivity to store depletion-induced apoptosis in neural cells. *J Biol Chem* 281: 37391–37403, 2006.
31. Chen YW, Chen SH, Chou W, Lo YM, Hung CH, and Lin MT. Exercise pretraining protects against cerebral ischaemia induced by heat stroke in rats. *Br J Sports Med* 41: 597–602, 2007.
32. Cheng B and Mattson MP. NT-3 and BDNF protect CNS neurons against metabolic/excitotoxic insults. *Brain Res* 640: 56–67, 1994.
33. Colcombe S and Kramer AF. Fitness effects on the cognitive function of older adults: a meta-analytic study. *Psychol Sci* 14: 125–130, 2003.
34. Colcombe SJ, Erickson KI, Scalf PE, Kim JS, Prakash R, McAuley E, Elavsky S, Marquez DX, Hu L, and Kramer AF. Aerobic exercise training increases brain volume in aging humans. *J Gerontol A Biol Sci Med Sci* 61: 1166–1170, 2006.
35. Costa DA, Cracchiolo JR, Bachstetter AD, *et al.* Enrichment improves cognition in AD mice by amyloid-related and unrelated mechanisms. *Neurobiol Aging* 28: 831–844, 2007.
- 35a. Cotman CW and Head E. The canine (dog) model of human aging and disease: dietary, environmental and immunotherapy approaches. *J Alzheimers Dis* 15: 685–707, 2008.
36. Cracchiolo JR, Mori T, Nazian SJ, Tan J, Potter H, and Arendash GW. Enhanced cognitive activity—over and above social or physical activity—is required to protect Alzheimer's mice against cognitive impairment, reduce abeta deposition, and increase synaptic immunoreactivity. *Neurobiol Learn Mem* 88: 277–294, 2007.
37. Cui L, Hofer T, Rani A, Leeuwenburgh C, and Foster TC. Comparison of lifelong and late life exercise on oxidative stress in the cerebellum. *Neurobiol Aging* 30: 903–909, 2009.
38. Dickey CA, Kamal A, Lundgren K, Klosak N, Bailey RM, Dunmore J, Ash P, Shoraka S, Zlatkovic J, Eckman CB, Patterson C, Dickson DW, Nahman NS Jr., Hutton M, Burrows F, and Petrucelli L. The high-affinity HSP90-CHIP complex recognizes and selectively degrades phosphorylated tau client proteins. *J Clin Invest* 117: 648–658, 2007.
39. Dietrich MO, Andrews ZB, and Horvath TL. Exercise-induced synaptogenesis in the hippocampus is dependent on UCP2-regulated mitochondrial adaptation. *J Neurosci* 28: 10766–10771, 2008.
40. Dong S, Li C, Wu P, Tsien JZ, and Hu Y. Environment enrichment rescues the neurodegenerative phenotypes in presenilins-deficient mice. *Eur J Neurosci* 26: 101–112, 2007.
41. Duan W, Guo Z, Jiang H, Ladenheim B, Xu X, Cadet JL, and Mattson MP. Paroxetine retards disease onset and progression in Huntington mutant mice. *Ann Neurol* 55: 590–594, 2004.
42. Duan W, Guo Z, Jiang H, Ware M, Li XJ, and Mattson MP. Dietary restriction normalizes glucose metabolism and BDNF levels, slows disease progression, and increases survival in huntingtin mutant mice. *Proc Natl Acad Sci U S A* 100: 2911–2916, 2003.
43. Duan W, Guo Z, Jiang H, Ware M, and Mattson MP. Reversal of behavioral and metabolic abnormalities, and insulin resistance syndrome, by dietary restriction in mice deficient in brain-derived neurotrophic factor. *Endocrinology* 144: 2446–2453, 2003.
44. Duan W and Mattson MP. Dietary restriction and 2-deoxyglucose administration improve behavioral outcome and reduce degeneration of dopaminergic neurons in models of Parkinson's disease. *J Neurosci Res* 57: 195–206, 1999.
- 44a. Duan W, Peng Q, Masuda N, Ford E, Tryggestad E, Ladenheim B, Zhao M, Cadet JL, Wong J, and Ross CA. Sertraline slows disease progression and increases neurogenesis in N171-82Q mouse model of Huntington's disease. *Neurobiol Dis* 30: 312–322, 2008.
45. Durany N, Michel T, Kurt J, Cruz-Sanchez FF, Cervos-Navarro J, and Riederer P. Brain-derived neurotrophic factor and neurotrophin-3 levels in Alzheimer's disease brains. *Int J Dev Neurosci* 18: 807–813, 2000.
46. Esposito L, Raber J, Kekonius L, Yan F, Yu GQ, Bien-Ly N, Puolivali J, Searce-Levie K, Masliah E, and Mucke L. Reduction in mitochondrial superoxide dismutase modulates Alzheimer's disease-like pathology and accelerates the onset of behavioral changes in human amyloid precursor protein transgenic mice. *J Neurosci* 26: 5167–5179, 2006.
47. Etgen T, Sander D, Huntgeburth U, Poppert H, Forstl H, and Bickel H. Physical activity and incident cognitive impairment in elderly persons: the INVADE study. *Arch Intern Med* 170: 186–193, 2010.
48. Evans CG, Wisén S, and Gestwicki JE. Heat shock proteins 70 and 90 inhibit early stages of amyloid beta-(1–42) aggregation *in vitro*. *J Biol Chem* 281: 33182–33191, 2006.
49. Farr SA, Yamada KA, Butterfield DA, *et al.* Obesity and hypertriglyceridemia produce cognitive impairment. *Endocrinology* 149: 2628–2636, 2008.
50. Ferris LT, Williams JS, and Shen CL. The effect of acute exercise on serum brain-derived neurotrophic factor levels and cognitive function. *Med Sci Sports Exerc* 39: 728–734, 2007.
51. Fontan-Lozano A, Saez-Cassanelli JL, Inda MC, *et al.* Caloric restriction increases learning consolidation and facilitates synaptic plasticity through mechanisms dependent on NR2B subunits of the NMDA receptor. *J Neurosci* 27: 10185–10195, 2007.
52. Fratiglioni L, Paillard-Borg S, and Winblad B. An active and socially integrated lifestyle in late life might protect against dementia. *Lancet Neurol* 3: 343–353, 2004.
53. Fratiglioni L and Wang HX. Brain reserve hypothesis in dementia. *J Alzheimers Dis* 12: 11–22, 2007.
54. Friedland RP, Fritsch T, Smyth KA, *et al.* Patients with Alzheimer's disease have reduced activities in midlife compared with healthy control-group members. *Proc Natl Acad Sci U S A* 98: 3440–3445, 2001.
55. Fujikake N, Nagai Y, Popiel HA, Okamoto Y, Yamaguchi M, and Toda T. Heat shock transcription factor 1-activating compounds suppress polyglutamine-induced neurodegen-



- eration through induction of multiple molecular chaperones. *J Biol Chem* 283: 26188–26197, 2008.
56. Golden E, Emiliano A, Maudsley S, Windham BG, Carlson OD, Egan JM, Driscoll I, Ferrucci L, Martin B, and Mattson MP. Circulating brain-derived neurotrophic factor and indices of metabolic and cardiovascular health: data from the Baltimore Longitudinal Study of Aging. *PLoS ONE* 5: e10099, 2010.
  57. Gomez-Pinilla F, Vaynman S, and Ying Z. Brain-derived neurotrophic factor functions as a metabotrophin to mediate the effects of exercise on cognition. *Eur J Neurosci* 28: 2278–2287, 2008.
  58. Gonzalez B, Hernandez R, and Manso R. Stress proteins of 70 kDa in chronically exercised skeletal muscle. *Pflugers Arch* 440: 42–49, 2000.
  59. Gorski JA, Balogh SA, Wehner JM, and Jones KR. Learning deficits in forebrain-restricted brain-derived neurotrophic factor mutant mice. *Neuroscience* 121: 341–354, 2003.
  60. Granholm AC, Bimonte-Nelson HA, Moore AB, Nelson ME, Freeman LR, and Sambamurti K. Effects of a saturated fat and high cholesterol diet on memory and hippocampal morphology in the middle-aged rat. *J Alzheimers Dis* 14: 133–145, 2008.
  61. Greenwood CE and Winocur G. Cognitive impairment in rats fed high-fat diets: a specific effect of saturated fatty-acid intake. *Behav Neurosci* 110: 451–459, 1996.
  - 61a. Grote HE, Bull ND, Howard ML, van Dellen A, Blake-more C, Bartlett PF, and Hannan AJ. Cognitive disorders and neurogenesis deficits in Huntington's disease mice are rescued by fluoxetine. *Eur J Neurosci* 22: 2081–2088, 2005.
  62. Gunstad J, Paul RH, Cohen RA, et al. Relationship between body mass index and brain volume in healthy adults. *Int J Neurosci* 118: 1582–1593, 2008.
  63. Gunstad J, Paul RH, Cohen RA, Tate DF, Spitznagel MB, and Gordon E. Elevated body mass index is associated with executive dysfunction in otherwise healthy adults. *Compr Psychiatry* 48: 57–61, 2007.
  64. Gunstad J, Schofield P, Paul RH, et al. BDNF Val66Met polymorphism is associated with body mass index in healthy adults. *Neuropsychobiology* 53: 153–156, 2006.
  65. Guo Q, Sebastian L, Sopher BL, Miller MW, Glazner GW, Ware CB, Martin GM, and Mattson MP. Neurotrophic factors [activity-dependent neurotrophic factor (ADNF) and basic fibroblast growth factor (bFGF)] interrupt excitotoxic neurodegenerative cascades promoted by a PS1 mutation. *Proc Natl Acad Sci U S A* 96: 4125–4130, 1999.
  66. Guo Z, Lee J, Lane M, and Mattson M. Iodoacetate protects hippocampal neurons against excitotoxic and oxidative injury: involvement of heat-shock proteins and Bcl-2. *J Neurochem* 79: 361–370, 2001.
  67. Halagappa VK, Guo Z, Pearson M, et al. Intermittent fasting and caloric restriction ameliorate age-related behavioral deficits in the triple-transgenic mouse model of Alzheimer's disease. *Neurobiol Dis* 26: 212–220, 2007.
  68. Harburger LL, Nzerem CK, and Frick KM. Single enrichment variables differentially reduce age-related memory decline in female mice. *Behav Neurosci* 121: 679–688, 2007.
  69. Hashimoto T and Watanabe S. Chronic food restriction enhances memory in mice—analysis with matched drive levels. *Neuroreport* 16: 1129–1133, 2005.
  70. Hellweg R, Lohmann P, Huber R, Kuhl A, and Riepe MW. Spatial navigation in complex and radial mazes in APP23 animals and neurotrophin signaling as a biological marker of early impairment. *Learn Mem* 13: 63–71, 2006.
  71. Herring A, Ambree O, Tomm M, et al. Environmental enrichment enhances cellular plasticity in transgenic mice with Alzheimer-like pathology. *Exp Neurol* 216: 184–192, 2009.
  72. Herring A, Blome M, Ambree O, Sachser N, Paulus W, and Keyvani K. Reduction of cerebral oxidative stress following environmental enrichment in mice with Alzheimer-like pathology. *Brain Pathol* 20: 166–175, 2010.
  73. Ho L, Qin W, Pompl PN, et al. Diet-induced insulin resistance promotes amyloidosis in a transgenic mouse model of Alzheimer's disease. *FASEB J* 18: 902–904, 2004.
  74. Holmes MM, Galea LA, Mistlberger RE, and Kempermann G. Adult hippocampal neurogenesis and voluntary running activity: circadian and dose-dependent effects. *J Neurosci Res* 76: 216–222, 2004.
  75. Honea RA, Thomas GP, Harsha A, et al. Cardiorespiratory fitness and preserved medial temporal lobe volume in Alzheimer disease. *Alzheimer Dis Assoc Disord* 23: 188–197, 2009.
  76. Hu YS, Xu P, Pigino G, Brady ST, Larson J, and Lazarov O. Complex environment experience rescues impaired neurogenesis, enhances synaptic plasticity, and attenuates neuropathology in familial Alzheimer's disease-linked APPswe/PS1{delta}E9 mice. *FASEB J* 24: 1667–1681, 2010.
  77. Hwang LL, Wang CH, Li TL, et al. Sex differences in high-fat diet-induced obesity, metabolic alterations and learning, and synaptic plasticity deficits in mice. *Obesity (Silver Spring)* 18: 463–469, 2010.
  78. Hyun DH, Emerson SS, Jo DG, Mattson MP, and de Cabo R. Calorie restriction up-regulates the plasma membrane redox system in brain cells and suppresses oxidative stress during aging. *Proc Natl Acad Sci U S A* 103: 19908–19912, 2006.
  79. Ickes BR, Pham TM, Sanders LA, Albeck DS, Mohammed AH, and Granholm AC. Long-term environmental enrichment leads to regional increases in neurotrophin levels in rat brain. *Exp Neurol* 164: 45–52, 2000.
  80. Jankowsky JL, Melnikova T, Fadale DJ, et al. Environmental enrichment mitigates cognitive deficits in a mouse model of Alzheimer's disease. *J Neurosci* 25: 5217–5224, 2005.
  81. Jankowsky JL, Xu G, Fromholt D, Gonzales V, and Borchelt DR. Environmental enrichment exacerbates amyloid plaque formation in a transgenic mouse model of Alzheimer disease. *J Neuropathol Exp Neurol* 62: 1220–1227, 2003.
  82. Jurdak N, Lichtenstein AH, and Kanarek RB. Diet-induced obesity and spatial cognition in young male rats. *Nutr Neurosci* 11: 48–54, 2008.
  83. Kakimura J, Kitamura Y, Takata K, Umeki M, Suzuki S, Shibagaki K, Taniguchi T, Nomura Y, Gebicke-Haerter PJ, Smith MA, Perry G, and Shimohama S. Microglial activation and amyloid-beta clearance induced by exogenous heat-shock proteins. *FASEB J* 16: 601–603, 2002.
  84. Katzman R, Terry R, DeTeresa R, et al. Clinical, pathological, and neurochemical changes in dementia: a subgroup with preserved mental status and numerous neocortical plaques. *Ann Neurol* 23: 138–144, 1988.
  85. Keller JN, Hanni KB, and Markesbery WR. Impaired proteasome function in Alzheimer's disease. *J Neurochem* 75: 436–439, 2000.
  86. Keller JN, Kindy MS, Holtsberg FW, St. Clair DK, Yen HC, Germeyer A, Steiner SM, Bruce-Keller AJ, Hutchins JB, and

- Mattson MP. Mitochondrial manganese superoxide dismutase prevents neural apoptosis and reduces ischemic brain injury: suppression of peroxynitrite production, lipid peroxidation, and mitochondrial dysfunction. *J Neurosci* 18: 687–697, 1998.
87. Kernie SG, Liebl DJ, and Parada LF. BDNF regulates eating behavior and locomotor activity in mice. *EMBO J* 19: 1290–1300, 2000.
  88. Koren J 3rd, Jinwal UK, Lee DC, Jones JR, Shults CL, Johnson AG, Anderson LJ, and Dickey CA. Chaperone signalling complexes in Alzheimer's disease. *J Cell Mol Med* 13: 619–630, 2009.
  89. Korte M, Carroll P, Wolf E, Brem G, Thoenen H, and Bonhoeffer T. Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc Natl Acad Sci U S A* 92: 8856–8860, 1995.
  90. Korte M, Staiger V, Griesbeck O, Thoenen H, and Bonhoeffer T. The involvement of brain-derived neurotrophic factor in hippocampal long-term potentiation revealed by gene targeting experiments. *J Physiol Paris* 90: 157–164, 1996.
  91. This reference has been deleted.
  92. Lafenetre P, Leske O, Ma-Hogemeie Z, et al. Exercise can rescue recognition memory impairment in a model with reduced adult hippocampal neurogenesis. *Front Behav Neurosci* 3: 34, 2010.
  93. Larson EB, Wang L, Bowen JD, et al. Exercise is associated with reduced risk for incident dementia among persons 65 years of age and older. *Ann Intern Med* 144: 73–81, 2006.
  94. Lazarov O, Mattson MP, Peterson DA, Pimplikar SW, and van Praag H. When neurogenesis encounters aging and disease. *Trends Neurosci* 33: 569–579, 2010.
  95. Lazarov O, Robinson J, Tang YP, et al. Environmental enrichment reduces abeta levels and amyloid deposition in transgenic mice. *Cell* 120: 701–713, 2005.
  96. Lee J, Bruce-Keller AJ, Kruman Y, Chan SL, and Mattson MP. 2-Deoxy-D-glucose protects hippocampal neurons against excitotoxic and oxidative injury: evidence for the involvement of stress proteins. *J Neurosci Res* 57: 48–61, 1999.
  97. Lee J, Duan W, and Mattson MP. Evidence that brain-derived neurotrophic factor is required for basal neurogenesis and mediates, in part, the enhancement of neurogenesis by dietary restriction in the hippocampus of adult mice. *J Neurochem* 82: 1367–1375, 2002.
  98. Lindqvist A, Mohapel P, Bouter B, et al. High-fat diet impairs hippocampal neurogenesis in male rats. *Eur J Neurol* 13: 1385–1388, 2006.
  99. Lindsay J, Laurin D, Verreault R, et al. Risk factors for Alzheimer's disease: a prospective analysis from the Canadian study of health and aging. *Am J Epidemiol* 156: 445–453, 2002.
  100. Lipsky RH, Xu K, Zhu D, et al. Nuclear factor kappaB is a critical determinant in N-methyl-D-aspartate receptor-mediated neuroprotection. *J Neurochem* 78: 254–264, 2001.
  101. Liu D, Chan SL, de Souza-Pinto NC, et al. Mitochondrial UCP4 mediates an adaptive shift in energy metabolism and increases the resistance of neurons to metabolic and oxidative stress. *Neuromol Med* 8: 389–414, 2006.
  102. Liu H, Wang H, Shenvi S, Hagen TM, and Liu RM. Glutathione metabolism during aging and in Alzheimer disease. *Ann N Y Acad Sci* 1019: 346–349, 2004.
  103. Lommatzsch M, Zingler D, Schuhbaeck K, et al. The impact of age, weight and gender on BDNF levels in human platelets and plasma. *Neurobiol Aging* 26: 115–123, 2005.
  104. Luchsinger JA, Tang MX, Shea S, and Mayeux R. Caloric intake and the risk of Alzheimer disease. *Arch Neurol* 59: 1258–1263, 2002.
  105. Martin B, Ji S, Maudsley S, and Mattson MP. "Control" laboratory rodents are metabolically morbid: why it matters. *Proc Natl Acad Sci U S A* 107: 6127–6133, 2010.
  106. Martin B, Mattson MP, and Maudsley S. Caloric restriction and intermittent fasting: two potential diets for successful brain aging. *Ageing Res Rev* 5: 332–353, 2006.
  107. Martin B, Pearson M, Brenneman R, Golden E, Keselman A, Iyun T, Carlson OD, Egan JM, Becker KG, Wood W 3rd, Prabhu V, de Cabo R, Maudsley S, and Mattson MP. Conserved and differential effects of dietary energy intake on the hippocampal transcriptomes of females and males. *PLoS ONE* 3: e2398, 2008.
  108. Maswood N, Young J, Tilmont E, Zhang Z, Gash DM, Gerhardt GA, Grondin R, Roth GS, Mattison J, Lane MA, Carson RE, Cohen RM, Mouton PR, Quigley C, Mattson MP, and Ingram DK. Caloric restriction increases neurotrophic factor levels and attenuates neurochemical and behavioral deficits in a primate model of Parkinson's disease. *Proc Natl Acad Sci U S A* 101: 18171–18176, 2004.
  109. Mattiasson G, Shamloo M, Gido G, Mathi K, Tomasevic G, Yi S, Warden CH, Castilho RF, Melcher T, Gonzalez-Zulueta M, Nikolich K, and Wieloch T. Uncoupling protein-2 prevents neuronal death and diminishes brain dysfunction after stroke and brain trauma. *Nat Med* 9: 1062–1068, 2003.
  110. Mattson MP. Hormesis defined. *Ageing Res Rev* 7: 1–7, 2008.
  111. Mattson MP. Metal-catalyzed disruption of membrane protein and lipid signaling in the pathogenesis of neurodegenerative disorders. *Ann N Y Acad Sci* 1012: 37–50, 2004.
  112. Mattson MP. Pathways towards and away from Alzheimer's disease. *Nature* 430: 631–639, 2004.
  113. Mattson MP. Roles of the lipid peroxidation product 4-hydroxynonenal in obesity, the metabolic syndrome, and associated vascular and neurodegenerative disorders. *Exp Gerontol* 44: 625–633, 2009.
  114. Mattson MP and Cheng A. Neurohormetic phytochemicals: low-dose toxins that induce adaptive neuronal stress responses. *Trends Neurosci* 29: 632–639, 2006.
  115. Mattson MP, Gleichmann M, and Cheng A. Mitochondria in neuroplasticity and neurological disorders. *Neuron* 60: 748–766, 2008.
  116. Mattson MP, Lovell MA, Furukawa K, and Markesbery WR. Neurotrophic factors attenuate glutamate-induced accumulation of peroxides, elevation of intracellular Ca<sup>2+</sup> concentration, and neurotoxicity and increase antioxidant enzyme activities in hippocampal neurons. *J Neurochem* 65: 1740–1751, 1995.
  117. Mattson MP and Magnus T. Ageing and neuronal vulnerability. *Nat Rev Neurosci* 7: 278–294, 2006.
  118. Mattson MP, Maudsley S, and Martin B. BDNF and 5-HT: a dynamic duo in age-related neuronal plasticity and neurodegenerative disorders. *Trends Neurosci* 27: 589–594, 2004.
  119. Monteleone P, Tortorella A, Martiadis V, Serritella C, Fuschino A, and Maj M. Opposite changes in the serum brain-derived neurotrophic factor in anorexia nervosa and obesity. *Psychosom Med* 66: 744–748, 2004.
  120. Morton JP, Kayani AC, McArdle A, and Drust B. The exercise-induced stress response of skeletal muscle, with specific emphasis on humans. *Sports Med* 39: 643–662, 2009.
  121. Murray AJ, Knight NS, Cochlin LE, et al. Deterioration of physical performance and cognitive function in rats

- with short-term high-fat feeding. *FASEB J* 23: 4353–4360, 2009.
122. Neeper SA, Gomez-Pinilla F, Choi J, and Cotman CW. Physical activity increases mRNA for brain-derived neurotrophic factor and nerve growth factor in rat brain. *Brain Res* 726: 49–56, 1996.
  123. Nelson RL, Guo Z, Halagappa VM, Pearson M, Gray AJ, Matsuoka Y, Brown M, Martin B, Iyun T, Maudsley S, Clark RF, and Mattson MP. Prophylactic treatment with paroxetine ameliorates behavioral deficits and retards the development of amyloid and tau pathologies in 3xTgAD mice. *Exp Neurol* 205: 166–176, 2007.
  124. Nichol K, Deeny SP, Seif J, Camaclang K, and Cotman CW. Exercise improves cognition and hippocampal plasticity in APOE epsilon4 mice. *Alzheimers Dement* 5: 287–294, 2009.
  125. Nichol KE, Parachikova AI, and Cotman CW. Three weeks of running wheel exposure improves cognitive performance in the aged Tg2576 mouse. *Behav Brain Res* 184: 124–132, 2007.
  126. Nichol KE, Poon WW, Parachikova AI, Cribbs DH, Glabe CG, and Cotman CW. Exercise alters the immune profile in Tg2576 Alzheimer mice toward a response coincident with improved cognitive performance and decreased amyloid. *J Neuroinflammation* 5: 13, 2008.
  127. Nithianantharajah J and Hannan AJ. The neurobiology of brain and cognitive reserve: mental and physical activity as modulators of brain disorders. *Prog Neurobiol* 89: 369–382, 2009.
  128. Nonomura T, Tsuchida A, Ono-Kishino M, Nakagawa T, Taiji M, and Noguchi H. Brain-derived neurotrophic factor regulates energy expenditure through the central nervous system in obese diabetic mice. *Int J Exp Diabetes Res* 2: 201–209, 2001.
  129. Parachikova A, Nichol KE, and Cotman CW. Short-term exercise in aged Tg2576 mice alters neuroinflammation and improves cognition. *Neurobiol Dis* 30: 121–129, 2008.
  130. Patterson SL, Abel T, Deuel TA, Martin KC, Rose JC, and Kandel ER. Recombinant BDNF rescues deficits in basal synaptic transmission and hippocampal LTP in BDNF knockout mice. *Neuron* 16: 1137–1145, 1996.
  131. Peila R, Rodriguez BL, and Launer LJ; Honolulu-Asia Aging Study. Type 2 diabetes, APOE gene, and the risk for dementia and related pathologies: the Honolulu-Asia aging study. *Diabetes* 51: 1256–1262, 2002.
  132. Pietropaolo S, Sun Y, Li R, Brana C, Feldon J, and Yee BK. The impact of voluntary exercise on mental health in rodents: a neuroplasticity perspective. *Behav Brain Res* 192: 42–60, 2008.
  133. Quartu M, Lai ML, and Del Fiacco M. Neurotrophin-like immunoreactivity in the human hippocampal formation. *Brain Res Bull* 48: 375–382, 1999.
  134. Radak Z, Kaneko T, Tahara S, et al. Regular exercise improves cognitive function and decreases oxidative damage in rat brain. *Neurochem Int* 38: 17–23, 2001.
  135. Rhodes JS, van Praag H, Jeffrey S, et al. Exercise increases hippocampal neurogenesis to high levels but does not improve spatial learning in mice bred for increased voluntary wheel running. *Behav Neurosci* 117: 1006–1016, 2003.
  136. Roe CM, Xiong C, Miller JP, and Morris JC. Education and Alzheimer disease without dementia: support for the cognitive reserve hypothesis. *Neurology* 68: 223–228, 2007.
  137. Rolland Y, Abellan van Kan G, and Vellas B. Physical activity and Alzheimer's disease: from prevention to therapeutic perspectives. *J Am Med Dir Assoc* 9: 390–405, 2008.
  138. Romanczyk TB, Weickert CS, Webster MJ, Herman MM, Akil M, and Kleinman JE. Alterations in trkB mRNA in the human prefrontal cortex throughout the lifespan. *Eur J Neurosci* 15: 269–280, 2002.
  139. Rossi C, Angelucci A, Costantin L, et al. Brain-derived neurotrophic factor (BDNF) is required for the enhancement of hippocampal neurogenesis following environmental enrichment. *Eur J Neurosci* 24: 1850–1856, 2006.
  140. Rovio S, Kareholt I, Helkala EL, et al. Leisure-time physical activity at midlife and the risk of dementia and Alzheimer's disease. *Lancet Neurol* 4: 705–711, 2005.
  141. Rovio S, Kareholt I, Viitanen M, et al. Work-related physical activity and the risk of dementia and Alzheimer's disease. *Int J Geriatr Psychiatry* 22: 874–882, 2007.
  142. Rupniak HT, Joy KA, Atkin C, Brown G, Barnes JC, Dectrow SR, Malfroy B, Wong T, Anderson IK, Molloy CR, Mills GI, and Soden P. Oxidative neuropathology and putative chemical entities for Alzheimer's disease: neuroprotective effects of salen-manganese catalytic anti-oxidants. *Neurotox Res* 2: 167–178, 2000.
  143. Sabia S, Kivimaki M, Shipley MJ, Marmot MG, and Singh-Manoux A. Body mass index over the adult life course and cognition in late midlife: the whitehall II cohort study. *Am J Clin Nutr* 89: 601–607, 2009.
  144. Scarmeas N, Stern Y, Mayeux R, Manly JJ, Schupf N, and Luchsinger JA. Mediterranean diet and mild cognitive impairment. *Arch Neurol* 66: 216–225, 2009.
  145. Schäbitz WR, Sommer C, Zoder W, Kiessling M, Schwabinger M, and Schwab S. Intravenous brain-derived neurotrophic factor reduces infarct size and counterregulates Bax and Bcl-2 expression after temporary focal cerebral ischemia. *Stroke* 31: 2212–2217, 2000.
  146. Schulte-Herbruggen O, Eckart S, Deicke U, et al. Age-dependent time course of cerebral brain-derived neurotrophic factor, nerve growth factor, and neurotrophin-3 in APP23 transgenic mice. *J Neurosci Res* 86: 2774–2783, 2008.
  147. Sharma S and Kaur G. Neuroprotective potential of dietary restriction against kainate-induced excitotoxicity in adult male Wistar rats. *Brain Res Bull* 67: 482–491, 2005.
  148. Shi L, Adams MM, Linville MC, et al. Caloric restriction eliminates the aging-related decline in NMDA and AMPA receptor subunits in the rat hippocampus and induces homeostasis. *Exp Neurol* 206: 70–79, 2007.
  149. Shieh PB and Ghosh A. Molecular mechanisms underlying activity-dependent regulation of BDNF expression. *J Neurobiol* 41: 127–134, 1999.
  150. Spina MB, Squinto SP, Miller J, Lindsay RM, and Hyman C. Brain-derived neurotrophic factor protects dopamine neurons against 6-hydroxydopamine and N-methyl-4-phenylpyridinium ion toxicity: involvement of the glutathione system. *J Neurochem* 59: 99–106, 1992.
  151. Stranahan AM, Arumugam TV, Cutler RG, Lee K, Egan JM, and Mattson MP. Diabetes impairs hippocampal function through glucocorticoid-mediated effects on new and mature neurons. *Nat Neurosci* 11: 309–317, 2008.
  152. Stranahan AM, Lee K, Becker KG, Zhang Y, Maudsley S, Martin B, Cutler RG, and Mattson MP. Hippocampal gene expression patterns underlying the enhancement of memory by running in aged mice. *Neurobiol Aging* 31: 1937–1949, 2010.
  153. Stranahan AM, Lee K, Martin B, et al. Voluntary exercise and caloric restriction enhance hippocampal dendritic



- spine density and BDNF levels in diabetic mice. *Hippocampus* 19: 951–961, 2009.
154. Stranahan AM, Norman ED, Lee K, *et al.* Diet-induced insulin resistance impairs hippocampal synaptic plasticity and cognition in middle-aged rats. *Hippocampus* 18: 1085–1088, 2008.
  155. Sultana R, Perluigi M, and Butterfield DA. Oxidatively modified proteins in Alzheimer's disease (AD), mild cognitive impairment and animal models of AD: role of Abeta in pathogenesis. *Acta Neuropathol* 118: 131–150, 2009.
  156. Tanaka J, Horiike Y, Matsuzaki M, Miyazaki T, Ellis-Davies GC, and Kasai H. Protein synthesis and neurotrophin-dependent structural plasticity of single dendritic spines. *Science* 319: 1683–1687, 2008.
  157. Tapia-Arancibia L, Aliaga E, Silhol M, and Arancibia S. New insights into brain BDNF function in normal aging and Alzheimer disease. *Brain Res Rev* 59: 201–220, 2008.
  158. Tapia-Arancibia L, Rage F, Givalois L, and Arancibia S. Physiology of BDNF: focus on hypothalamic function. *Front Neuroendocrinol* 25: 77–107, 2004.
  159. Tonra JR, Ono M, Liu X, *et al.* Brain-derived neurotrophic factor improves blood glucose control and alleviates fasting hyperglycemia in C57BLKS-lepr(db)/lepr(db) mice. *Diabetes* 48: 588–594, 1999.
  160. Tozuka Y, Wada E, and Wada K. Diet-induced obesity in female mice leads to peroxidized lipid accumulations and impairment of hippocampal neurogenesis during the early life of their offspring. *FASEB J* 23: 1920–1934, 2009.
  161. Um HS, Kang EB, Leem YH, *et al.* Exercise training acts as a therapeutic strategy for reduction of the pathogenic phenotypes for Alzheimer's disease in an NSE/APPsw-transgenic model. *Int J Mol Med* 22: 529–539, 2008.
  162. Valenzuela M and Sachdev P. Can cognitive exercise prevent the onset of dementia? Systematic review of randomized clinical trials with longitudinal follow-up. *Am J Geriatr Psychiatry* 17: 179–187, 2009.
  163. Valenzuela MJ. Brain reserve and the prevention of dementia. *Curr Opin Psychiatry* 21: 296–302, 2008.
  164. Valenzuela MJ, Sachdev P, Wen W, Chen X, and Brodaty H. Lifespan mental activity predicts diminished rate of hippocampal atrophy. *PLoS ONE* 3: e2598, 2008.
  165. van Praag H. Exercise and the brain: something to chew on. *Trends Neurosci* 32: 283–290, 2009.
  166. van Praag H, Kempermann G, and Gage FH. Neural consequences of environmental enrichment. *Nat Rev Neurosci* 1: 191–198, 2000.
  167. van Praag H, Kempermann G, and Gage FH. Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat Neurosci* 2: 266–270, 1999.
  168. Vargas MR and Johnson JA. The Nrf2-ARE cytoprotective pathway in astrocytes. *Expert Rev Mol Med* 11: e17, 2009.
  169. Vasuta C, Caunt C, James R, *et al.* Effects of exercise on NMDA receptor subunit contributions to bidirectional synaptic plasticity in the mouse dentate gyrus. *Hippocampus* 17: 1201–1208, 2007.
  170. Vaynman S, Ying Z, and Gomez-Pinilla F. Hippocampal BDNF mediates the efficacy of exercise on synaptic plasticity and cognition. *Eur J Neurosci* 20: 2580–2590, 2004.
  171. White CL, Pistell PJ, Purpera MN, *et al.* Effects of high fat diet on morris maze performance, oxidative stress, and inflammation in rats: contributions of maternal diet. *Neurobiol Dis* 35: 3–13, 2009.
  172. Williams BJ, Eriksdotter-Jonhagen M, and Granholm AC. Nerve growth factor in treatment and pathogenesis of Alzheimer's disease. *Prog Neurobiol* 80: 114–128, 2006.
  173. Wilson RS, Krueger KR, Arnold SE, *et al.* Loneliness and risk of Alzheimer disease. *Arch Gen Psychiatry* 64: 234–240, 2007.
  174. Wilson RS, Scherr PA, Schneider JA, Tang Y, and Bennett DA. Relation of cognitive activity to risk of developing Alzheimer disease. *Neurology* 69: 1911–1920, 2007.
  175. Wolf SA, Kronenberg G, Lehmann K, *et al.* Cognitive and physical activity differently modulate disease progression in the amyloid precursor protein (APP)-23 model of Alzheimer's disease. *Biol Psychiatry* 60: 1314–1323, 2006.
  176. Wu A, Ying Z, and Gomez-Pinilla F. The interplay between oxidative stress and brain-derived neurotrophic factor modulates the outcome of a saturated fat diet on synaptic plasticity and cognition. *Eur J Neurosci* 19: 1699–1707, 2004.
  177. Xu W, Qiu C, Winblad B, and Fratiglioni L. The effect of borderline diabetes on the risk of dementia and Alzheimer's disease. *Diabetes* 56: 211–216, 2007.
  178. Xu X, Zhan M, Duan W, Prabhu V, Brenneman R, Wood W, Firman J, Li H, Zhang P, Ibe C, Zonderman AB, Longo DL, Poosala S, Becker KG, and Mattson MP. Gene expression atlas of the mouse central nervous system: impact and interactions of age, energy intake and gender. *Genome Biol* 8: R234, 2007.
  179. Yamanaka M, Itakura Y, Tsuchida A, Nakagawa T, and Taiji M. Brain-derived neurotrophic factor (BDNF) prevents the development of diabetes in prediabetic mice. *Biomed Res* 29: 147–153, 2008.
  180. Yanai S, Okaichi Y, and Okaichi H. Long-term dietary restriction causes negative effects on cognitive functions in rats. *Neurobiol Aging* 25: 325–332, 2004.
  181. Yu H, Bi Y, Ma W, *et al.* Long-term effects of high lipid and high energy diet on serum lipid, brain fatty acid composition, and memory and learning ability in mice. *Int J Dev Neurosci* 28: 271–276, 2010.
  182. Yu ZF and Mattson MP. Dietary restriction and 2-deoxyglucose administration reduce focal ischemic brain damage and improve behavioral outcome: evidence for a preconditioning mechanism. *J Neurosci Res* 57: 830–839, 1999.
  183. Yuede CM, Zimmerman SD, Dong H, *et al.* Effects of voluntary and forced exercise on plaque deposition, hippocampal volume, and behavior in the Tg2576 mouse model of Alzheimer's disease. *Neurobiol Dis* 35: 426–432, 2009.
  184. Zahn JM, Poosala S, Owen AB, Ingram DK, Lustig A, Carter A, Weeraratna AT, Taub DD, Gorospe M, Mazan-Mamczarz K, Lakatta EG, Boheler KR, Xu X, Mattson MP, Falco G, Ko MS, Schlessinger D, Firman J, Kummerfeld SK, Wood WH 3rd, Zonderman AB, Kim SK, and Becker KG. AGEMAP: a gene expression database for aging in mice. *PLoS Genet* 3: e201, 2007.
  185. Zhu SW, Codita A, Bogdanovic N, Hjerling-Leffler J, Ernfors P, Winblad B, Dickins DW, and Mohammed AH. Influence of environmental manipulation on exploratory behaviour in male BDNF knockout mice. *Behav Brain Res* 197: 339–346, 2009.
  186. Zhu SW, Yee BK, Nyffeler M, Winblad B, Feldon J, and Mohammed AH. Influence of differential housing on emotional behaviour and neurotrophin levels in mice. *Behav Brain Res* 169: 10–20, 2006.

187. Zhu X, Raina AK, Lee HG, Casadesus G, Smith MA, and Perry G. Oxidative stress signalling in Alzheimer's disease. *Brain Res* 1000: 32–39, 2004.
188. Ziegenhorn AA, Schulte-Herbruggen O, Danker-Hopfe H, *et al.* Serum neurotrophins—a study on the time course and influencing factors in a large old age sample. *Neurobiol Aging* 28: 1436–1445, 2007.

Address correspondence to:

Dr. Mark P. Mattson

Laboratory of Neurosciences

National Institute of Aging Intramural Research Program  
Baltimore, MD 21224

E-mail: mattsonm@grc.nia.nih.gov

Date of first submission to ARS Central, August 30, 2010; date of acceptance, September 19, 2010.

#### Abbreviations Used

A $\beta$  = amyloid  $\beta$ -peptide  
ACSRP = adaptive cellular stress response pathways  
AD = Alzheimer's disease  
AICD = intracellular amyloid precursor protein domain  
APP = amyloid precursor protein  
BDNF = brain-derived neurotrophic factor

BMI = body mass index  
cGMP = cyclic guanosine monophosphate  
CR = caloric restriction  
CREB = cyclic AMP response element binding protein  
EE = environmental enrichment  
ER = endoplasmic reticulum  
FGF2 = fibroblast growth factor 2  
GRP78 = glucose-regulated protein 78  
HD = Huntington's disease  
HSP70 = heat-shock protein 70  
Htt = huntingtin  
IF = intermittent fasting  
IP<sub>3</sub>R = IP<sub>3</sub> receptor  
LTP = long-term potentiation  
MAOS = membrane-associated oxidative stress  
NFT = neurofibrillary tangles  
NGF = nerve growth factor  
PD = Parkinson's disease  
PPases = protein phosphatases  
PS1 = presenilin-1  
PtdS = phosphatidylserine  
ROS = reactive oxygen species  
RyR = ryanodine receptor  
SERCA = smooth endoplasmic reticulum Ca<sup>2+</sup>-ATPase  
SOD = superoxide dismutase  
UCP = uncoupling protein  
VDCC = voltage-dependent Ca<sup>2+</sup> channels

**This article has been cited by:**

1. Ajay S. Unnithan, Hailey J.H. Choi, Amanda M. Titler, Jessica M. Posimo, Rehana K. Leak. 2012. Rescue from a two hit, high-throughput model of neurodegeneration with N-acetyl cysteine. *Neurochemistry International* **61**:3, 356-368. [[CrossRef](#)]
2. Daniela Hartl, Victoria Schuldt, Stephanie Forler, Claus Zabel, Joachim Klose, Michael Rohe. 2012. Presymptomatic Alterations in Energy Metabolism and Oxidative Stress in the APP23 Mouse Model of Alzheimer Disease. *Journal of Proteome Research* 120522153528002. [[CrossRef](#)]
3. Mark P. Mattson, Nicolas G. Bazan Apoptosis and Necrosis 663-676. [[CrossRef](#)]
4. Giles E. Hardingham , Stuart A. Lipton . 2011. Regulation of Neuronal Oxidative and Nitrosative Stress by Endogenous Protective Pathways and Disease Processes. *Antioxidants & Redox Signaling* **14**:8, 1421-1424. [[Abstract](#)] [[Full Text HTML](#)] [[Full Text PDF](#)] [[Full Text PDF with Links](#)]
5. Ailton Melo, Larissa Monteiro, Rute M. F. Lima, Diêgo M. de Oliveira, Martins D. de Cerqueira, Ramon S. El-Bachá. 2011. Oxidative Stress in Neurodegenerative Diseases: Mechanisms and Therapeutic Perspectives. *Oxidative Medicine and Cellular Longevity* **2011**, 1-14. [[CrossRef](#)]